

Rapid Quantification of the Activating Effects of Hydrogen-Bonding Catalysts with a Colorimetric Sensor

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Supporting Information

ABSTRACT: A sensor has been developed to quickly and simply assess the relative reactivity of different hydrogenbonding catalysts. Specifically, blue-shifts seen upon treatment of H-bonding catalysts with the colorimetric compound 7-methyl-2-phenylimidazo[1,2-*a*]pyrazin-3(7H)-one correlate well to the K_{eq} of binding to the sensor. The blue-shifts also show a high degree of correlation with relative rates in Diels–Alder reactions of methyl vinyl ketone and cyclopentadiene employing the H-bonding catalysts. The relevance of the sensor blueshifts to the LUMO-lowering abilities of the H-bonding catalysts is discussed.

lectrophile activation by small-molecule hydrogen-bond donors has emerged as an important paradigm for enantioselective catalysis.¹ Nonetheless, a thorough understanding of the principles and features that govern the reactivity and selectivity of these catalysts remains incomplete. A number of physical organic measurements have provided scales that can be used to estimate the reactivity, such as pK_a tables,² nucleophilicity and electrophilicity parameters,³ Irving-Williams order,^{4,5} etc., but no scales have been made for all categories of hydrogen-bonding catalysts. Contributing to this problem is the large range of H-bond strengths, from 0.2 to 40 kcal/mol.⁶ While the strength of a H-bonding interaction can be inferred from $\Delta p K_{av}^{7,8}$ such a measurement gives an incomplete account with respect to catalysis since a water molecule poorly mimics a substrate. As a result, secondary interactions, such as sterics, dual H-bonding,9 and H-bonding directionality, between a H-bond donor and an electrophilic substrate are not fully incorporated. Here, we present a simple spectroscopic measurement using a colorimetric sensor to determine the effectiveness of H-bonding catalysts in electrophilic activation of a monodentate substrate. The measurement is effective for a range of catalysts encompassing a pK_a window of ∼7−20.

We assessed a number of methods to judge the ability of different H-bond donors to activate a carbonyl (LUMO-lowering) but found that methods effective for strong Lewis acids, such as changes in IR or NMR signals, provided insufficient signal or were technically challenging. In search of a simple, easily applied measurement, we elected to use a colorimetric sensor molecule. 7-Methyl-2-phenylimidazo[1,2-a]pyrazin-3(7H)-one (1), which gives good correlations between λ_{max} -shifts and the Fukuzumi parameters for a small

number of Lewis acids, $^{10-12}$ was discovered to give a readily discernible signal upon coordination (eq 1) with a range of H-



bond donors (Chart 1). Figure 1 illustrates the simplicity of the method, with changes in color that are readily visible to the naked eye upon saturation with different H-bonding catalysts.







Figure 1. Change in color upon addition of hydrogen-bonding catalysts (see Chart 1) to the pyrazinone sensor 1 in dichloromethane.

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Figure 2. Response of sensor 1 at 2.22×10^{-5} M to increasing amounts of N_rN' -di(3,5-bis(trifluoromethyl)phenyl)thiourea (4) in dichloromethane.

Table 1. Hydrogen-Bonding Catalyst Saturated λ_{max} and K_{eq} Values for Binding to 1, along with k_{cat} Values for the Reaction in Eq 2 at 1 mol% Catalyst Loading in Benzene

H-bond catalyst	pK _a (in DMSO)	$\lambda_{\max} \ (nm)$	$K_{\rm eq}~({ m M}^{-1})$	$k_{\rm cat}~({\rm s}^{-1})$
none		499		_ ^a
2	13.4 (ref 7)	490	1.67×10	1.26×10^{-6}
3	17.1 (ref 15) ^b	487	3.23×10	1.80×10^{-6}
4	8.5 (ref 7)	477	1.77×10^{3}	2.09×10^{-5}
5	12.8-13.6 (ref 16)	473	3.34×10^{3}	4.90×10^{-5}
6	12.8–13.6 (ref 16) ^c	465	3.47×10^{5}	1.79×10^{-4}

 ${}^{a}k_{uncat} = 7.50 \times 10^{-5} \text{ s}^{-1}$. ^bFor 2-naphthol. ^cFirst pK_a may be 1–2 units lower due to dicationic nature of **6**.



Figure 3. Correlation between wavelength-shift and K_{eq} .

Figure 2 further illustrates the blue-shift in the λ_{max} of the sensor when combined with increasing amounts of a Hbonding catalyst, in this case N,N'-di(3,5-bis(trifluoromethyl)phenyl)thiourea (4). With these data, K_{eq} values (Table 1) for the sensor-H-bond donor association¹³ could be readily obtained from the corresponding titration curves as illustrated for 4.¹⁴ The inverse of the λ_{max} -shift obtained upon saturation with **2**-6 showed a strong correlation with the K_{eq} value (Figure 3), indicating that this λ_{max} -shift could be used as a reliable indicator of the association between the sensor and a prospective H-bonding catalyst.

Importantly, this sensor coordinates very weakly to water ($\Delta \lambda_{max}$ at saturation = 3.4 nm), which is easily displaced by catalyst. Thus, implementation is simple: sufficient catalyst is

added until no further blue-shift is seen. At this point, any water has been displaced, and the sensor is saturated. The $\lambda_{\rm max}$ obtained at this juncture is then used in the correlations to binding $(K_{\rm eq})$ and rate $(k_{\rm rel})$ see below). For example, a measurement can be made using 10 μ g of the sensor and \leq 10 mg of the catalyst without special precautions to exclude moisture.

Diels–Alder reactions of $\alpha_{,\beta}$ -unsaturated carbonyl dienophiles are well established to undergo rate acceleration with Lewis acids by LUMO-lowering of the dienophile,^{17–20} and a similar activation is believed to operate for H-bonding catalysts.²¹ To limit the number of different interactions between the substrates and the H-bonding catalyst, the monodentate substrate methyl vinyl ketone was selected along with a nonbonding diene, cyclopentadiene (eq 2). Rate measurements by NMR^{22,23} showed a range of activities for different H-bonding catalysts (Table 1).

A plot of $\ln(k_{\rm rel})$ ($k_{\rm rel} = k_{\rm cat}/k_{\rm uncat}$) vs the inverse of the $\lambda_{\rm max}$ -shift (Figure 4) showed a strong correlation, indicating that the binding to the sensor provides a reasonable account of the LUMO-lowering ability of different H-bonding catalysts. In contrast, the p $K_{\rm a}$ values do not track well with the reactivity (Table 1, p $K_{\rm a}$ vs $k_{\rm cat}$).



Figure 4. Correlation of Diels–Alder $k_{\rm rel}$ values from different hydrogen-bonding catalysts with the wavelength-shifts of sensor **1**.

In conclusion, pyrazinone sensor 1 was found to rapidly provide a read-out of the relative reactivity of hydrogenbonding catalysts in the Diels—Alder reaction of methyl vinyl ketone and cyclopentadiene. Namely, catalysts that cause a greater blue-shift at saturation of the sensor are more reactive. Thus, it appears that the interaction between hydrogen-bond donors and the carbonyl of the sensor provides a good approximation of the LUMO-lowering potential available via Hbonding. These preliminary results support the use of sensor 1 as a tool to gauge the relatively reactivity of new H-bonding catalysts and to further the understanding of why some Hbonding catalysts are more effective than others. Exploration of additional H-bonding donors and Lewis acids with the pyrazinone sensor and with other reactions is underway.

ASSOCIATED CONTENT

S Supporting Information

Experimental procedures, kinetics results, and spectral data. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) For reviews, see: (a) Taylor, M. S.; Jacobsen, E. N. Angew. Chem, Int. Ed. 2006, 45, 1520–1543. (b) Doyle, A. G.; Jacobsen, E. N. Chem. Rev. 2007, 107, 5713–5743. (c) Connon, S. J. Chem.—Eur. J. 2006, 12, 5418–5427. (d) Connon, S. J. Angew. Chem., Int. Ed. 2006, 45, 3909–3912. (e) Connon, S. J. Chem. Commun. 2008, 2499–2510. (f) Pihko, P. M. Angew. Chem., Int. Ed. 2004, 43, 2062–2064. (g) Ting, A.; Schaus, S. E. Eur. J. Org. Chem. 2007, 5797–5949. (h) Akiyama, T.; Itoh, J.; Fuchibe, K. Adv. Synth. Catal. 2006, 348, 999–1010. (i) Akiyama, T. Chem. Rev. 2007, 107, 5744–5758. (j) Nagasawa, K.; Sohtome, Y. Synlett 2010, 000A–000V. (k) Schenker, S.; Zamfir, A.; Freund, M.; Tsogoeva, S. B. Eur. J. Org. Chem. 2011, 2209–2222. (l) Jensen, K. H.; Sigman, M. S. J. Org. Chem. 2010, 75, 7194–7201.

(2) Bordwell, F. G. Acc. Chem. Res. 1988, 21, 456-463.

(3) Mayr, H.; Kempf, B.; Ofial, A. R. Acc. Chem. Res. 2003, 36, 66– 77.

(4) (a) Irving, H.; Williams, R. J. P. Nature 1948, 162, 746–747.
(b) Irving, H.; Williams, R. J. P. J. Chem. Soc. 1953, 3192–3210.

(5) Examples of applications of Irving–Williams order: (a) Evans, D.

A.; Rovis, T.; Johnson, J. S. Pure Appl. Chem. 1999, 71, 1407-1415.

(b) Johnson, J. S.; Evans, D. A. Acc. Chem. Res. 2000, 33, 325-335.

(6) Steiner, T. Angew. Chem., Int. Ed. 2002, 41, 48-76.

(7) Jakab, G.; Tancon, C.; Zhang, Z.; Lippert, K. M.; Schreiner, P. R. Org. Lett. **2012**, *14*, 1724–1727.

(8) Gilli, P.; Pretto, L.; Bertolasi, V.; Gilli, G. Acc. Chem. Res. 2009, 42, 33-44.

(9) Annamalai, V. R.; Linton, E. C.; Kozlowski, M. C. Org. Lett. 2009, 11, 621–624.

(10) Takamuki, Y; Maki, S.; Niwa, H.; Ikeda, H.; Hirano, T. *Tetrahedron* **2005**, *61*, 10073–10080.

(11) Fukuzumi, S.; Kei Ohkubo, K. Chem.—Eur. J. 2000, 6, 4532–4535.

(12) Fukuzumi, S.; Kei Ohkubo, K. J. Am. Chem. Soc. 2002, 124, 10270-10271.

(13) Protonation of the pyrazinone sensor with methanesulfonic acid in DMSO has been reported: Nakai, S.; Yasui, M.; Nakazato, M.; Iwasaki, F.; Maki, S.; Niwa, H.; Ohashi, M.; Hirano, T. Bull. Chim. Soc. Jpn. 2003, 76, 2361–2387. Based on the K value of protonation, the sensor can be determined to have a corresponding pK of -1.0(DMSO). These data support interactions between the sensor and Hbond donors that do not involve proton transfer.

(14) See Supporting Information.

(15) Bordwell, F. G. Acc. Chem. Res. 1988, 21, 456-463.

(16) Koppel, I.; Koppel, J.; Leito, I.; Grehn, L. J. Phys. Org. Chem. 1996, 9, 265-268.

(17) Corey, E. J.; Loh, T.-P; Sarshar, S.; Azimioara, M. Tetrahedron Lett. **1992**, 33, 6945–6948.

(18) Goering, H. L.; Chang, C. J. Org. Chem. 1975, 11, 2565.

(19) Alston, P. V; Ottenbrite, R. M. J. Org. Chem. 1975, 8, 1111-1116.

(20) Williamson, K. L.; Hsu, Y. L. J. Am. Chem. Soc. 1970, 92, 7385–7389.

(21) Huang, Y.; Unni, A. K.; Thadani, A. N.; Rawal, V. H. Nature 2003, 424, 146.

(22) Schreiner, P. R. Chem. Soc. Rev. 2003, 32, 289-296.

(23) Wittkopp, A.; Schreiner, P. R. Chem.-Eur. J. 2003, 9, 407-414.